

# **qPCR Monitoring Data and the Proposed Criteria Revisions**

## ***One Local Lab's Journey Towards Implementation***

**Great Lakes Beach Association Conference  
October 16, 2012  
Julie Kinzelman, PhD  
City of Racine Health Department**

# Proposed US Standards – Freshwater

- **Health effect data correlated to culture (*E. coli* and enterococci) and qPCR (enterococci)**
- **Culture**
  - *E. coli*
    - GM of 126 cfu per 100 mL (30 – 90 days)
    - STV of 235 cfu per 100 mL measured using EPA Method 1603, or any other equivalent method that measures culturable *E. coli* (i.e. IDEXX or similar)
  - Enterococci
    - GM of 33 cfu per 100 mL (30 – 90 days)
    - STV of 61 cfu per 100 mL measured using EPA Method 1600
- **qPCR**
  - Enterococci using EPA Method A
    - GM of 475 CCE per 100 mL
    - STV criterion of 1,000 CCE per 100 mL

# Alternative Method/Indicator Combinations

- *5.2.3 Alternative Criteria Based on Novel Indicators or New Analytical Methods, without Site-Specific Epidemiological Studies*
  - *E. coli* by qPCR, for example
  - Approval if scientifically defensible and protective of the recreational use
  - Consistent and predictable site-specific performance
  - Describes a common level of water quality
  - Consistent over a recreational season for both EPA-approved and proposed alternative indicator-method combination.

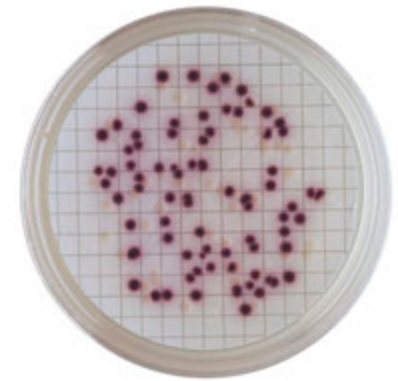
**Way back in 2005 Racine was  
thinking about rapid methods...**

*qPCR was under consideration for  
environmental use and also making it's  
appearance in the clinical arena*

**WE HAD QUESTIONS...**

# Remember QPCR is *not* Measuring the Same Thing as a Culture...

- QPCR differs from traditional culture-based assays in that it measures all DNA:
  - live cells
  - dead cells
  - non-culturable cells
  - free DNA
- Culture assays only measure cells possessing the ability to grow on the selective media you are using



# Correlation to Current Methods

- Do QPCR CCE correlate to currently approved methods?
  - Beach status (open/advisory/closed)
  - Actual counts
  - Is there diurnal variation and would it preclude early morning sampling?
  - Could we still composite sample?

# Composite Sampling

- Physical combining of multiple sub-samples into a single sample
  - Is it valid to composite?
  - Does the composite value fall within the range of individual values?
  - Would a composite sample mask a true elevation?
- Increases spatial coverage
- Maintains analytical costs
  - May be particularly useful when considering qPCR

# qPCR, 2005 – 2006

- Acquired Cepheid Smart Cycler using county CRI and PHEP grant funds
  - Increase public health capacity for molecular testing
  - Ability to accept routine testing as overflow site
- Trialed US EPA Taqman *Enterococcus* assay (pre Method A) vs. Method 1600
  - Composite sample analysis
    - North Beach
    - 4 transects
    - 12, 24, 36, and 48 inch depth zones
- Initial EPA validation study participant (2006)

# 2005, QPCR - Mean vs. Composite

Depth Zone	QPCR Mean	QPCR Composite	Method 1600 Mean	Method 1600 Composite
12"	121.48	156.94	35.6	56
24"	64.42	74.54	12.3	3
36"	14.17	5	6.2	10
48"	23.25	32.36	5.5	19

## QPCR

all NB comp:	Comp= 5	Mean= 67
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## CULTURE

all NB comp:	Comp = 37	Mean = 22
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**Initial Results Looked Promising**

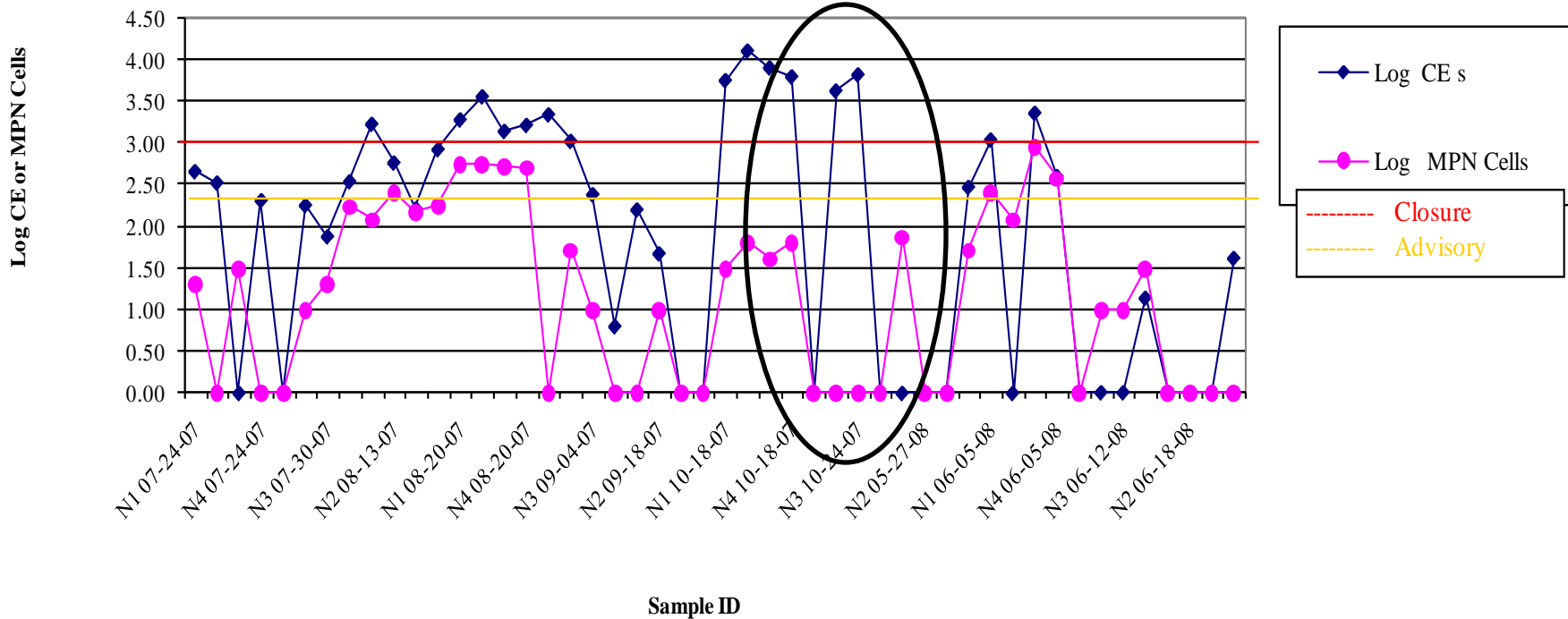
**We Decided to Do Some Comparative  
Testing in Conjunction with Routine  
Regulatory Monitoring**

# Ambient or Environmental DNA

- Does ambient background DNA exist?
- Is it in sufficient quantity to interfere with regulatory results?
- Can we determine when these conditions are likely to occur?
- Are they associated with environmental conditions?
  - Precipitation, wave height, turbidity
- Can they be corrected for?

# Uncorrected for Background DNA

North Beach 36": *E. coli* QPCR versus Colilert-18





# Correlation to "Beach Status" – *E. coli* (2007 – 2008)

Sample ID	Log CE s	Log MPN Cells	Corrected CE	Sample ID	Log CE s	Log MPN Cells	Corrected CE
N1 07-24-07	2.66	1.30	1.94	N1 10-18-07	3.44	1.49	2.72
N2 07-24-07	2.53	0.00	1.81	N2 10-18-07	3.47	1.80	2.75
N3 07-24-07	0.00	1.49		N3 10-18-07	3.33	1.61	2.61
N4 07-24-07	2.32	0.00	2.32	N4 10-18-07	3.37	1.80	2.65
N1 07-30-07	0.00	0.00	0.00	N1 10-24-07	0.00	0.00	0.00
N2 07-30-07	2.26	1.00	2.26	N2 10-24-07	2.70	0.00	1.98
N3 07-30-07	1.88	1.30	1.88	N3 10-24-07	2.86	0.00	2.14
N4 07-30-07	2.54	2.24	1.82	N4 10-24-07	0.00	0.00	0.00
N1 08-13-07	3.05	2.08	2.23	N1 05-27-08	0.00	1.87	0.00
N2 08-13-07	2.77	2.41	2.77	N2 05-27-08	0.00	0.00	0.00
N3 08-13-07	2.22	2.16	2.22	N3 05-27-08	0.00	0.00	0.00
N4 08-13-07	2.93	2.24	2.21	N4 05-27-08	2.48	1.72	1.76
N1 08-20-07	3.29	2.75	2.57	N1 06-05-08	3.18	2.41	2.46
N2 08-20-07	3.56	2.75	2.84	N2 06-05-08	0.00	2.08	-0.72
N3 08-20-07	3.15	2.72	2.43	N3 06-05-08	3.37	2.96	2.65
N4 08-20-07	3.22	2.70	2.50	N4 06-05-08	2.61	2.58	2.61
N1 09-04-07	3.05	1.00	2.33	N1 06-12-08	0.01	0.00	0.01
N2 09-04-07	3.03	1.72	2.31	N2 06-12-08	0.01	1.00	0.01
N3 09-04-07	2.39	1.00	1.67	N3 06-12-08	0.01	1.00	0.01
N4 09-04-07	0.80	0.00	0.80	N4 06-12-08	1.14	1.49	1.14
N1 09-18-07	2.20	0.00	2.20	N1 06-18-08	0.00	0.00	0.00
N2 09-18-07	1.68	1.00	1.68	N2 06-18-08	0.00	0.00	0.00
N3 09-18-07	0.00	0.00	0.00	N3 06-18-08	0.00	0.00	0.00
N4 09-18-07	0.00	0.00	0.00	N4 06-18-08	1.62	0.00	1.62

# Numerical/Regulatory Agreement – *E. coli* (2008)

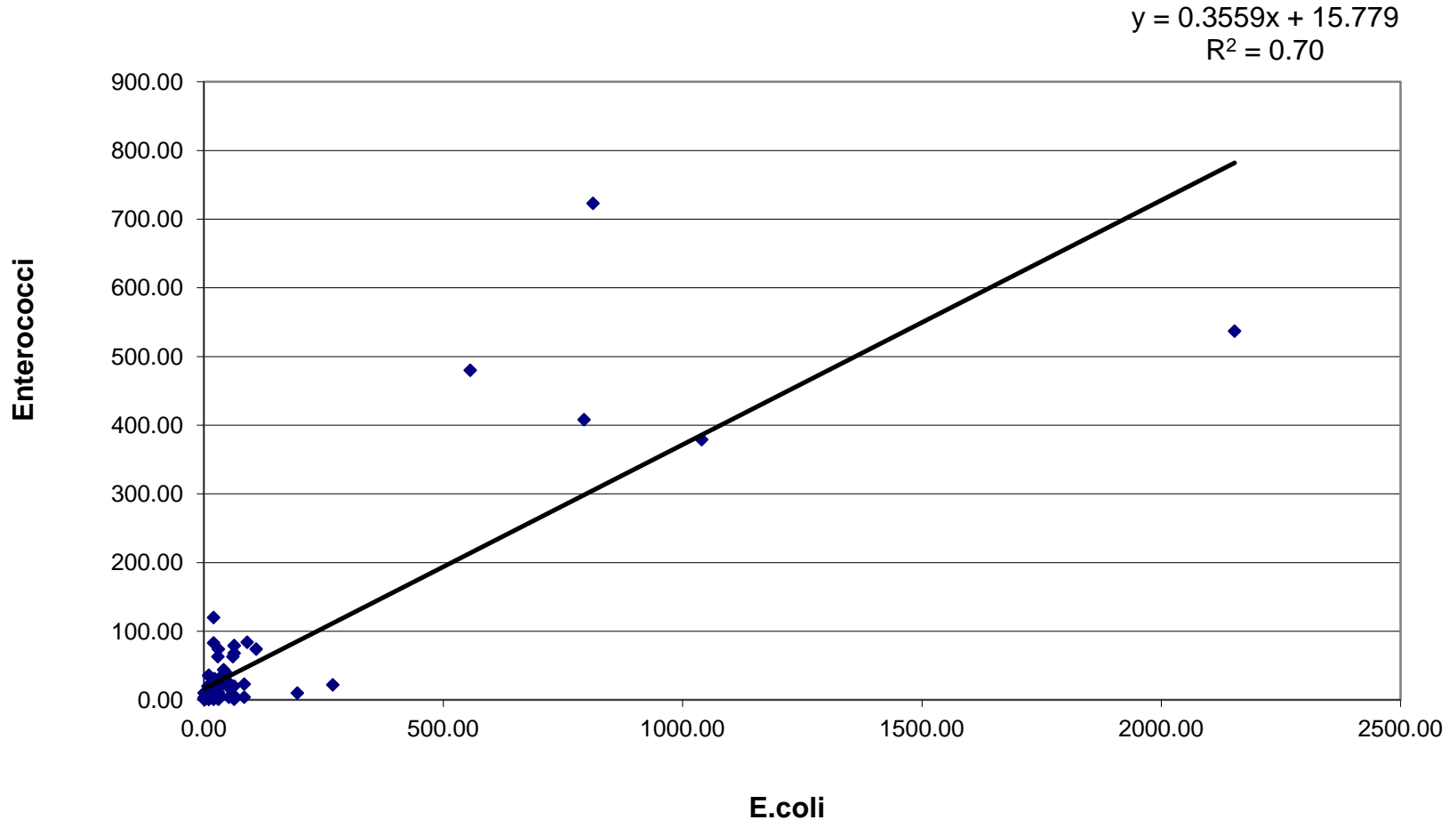
2008 <i>E. coli</i> , QPCR vs. Colilert-18 - Composite Samples, 1.0 m								
Date	QPCR Prediction (Status, CE)				Colilert-18 (Status, MPN)			
	North		Zoo		North		Zoo	
6/30/2008	open	10-30	open	<10	open	41	open	10
7/1/2008	open	20-50	open	10-30	open	10	open	10
7/2/2008	open	<10-10	open	<10-10	open	<10	open	31
7/3/2008	open	50-75	open	<10-20	open	213	advisory	419
7/7/2008	open	<10-20	open	<10-35	open	20	open	20
7/8/2008	open	<10-20	open	20-50	open	10	open	41
7/9/2008	open	<10-30	open	<10-20	open	<10	open	<10
7/10/2008	open	<10-20	open	<10-20	open	63	open	<10
7/11/2008	open	10-50	open	<10-30	open	20	open	41
7/14/2008	open	<10-20	open	<10-30	open	<10	open	<10
7/15/2008	open	10-30	open	<10-30	open	10	open	10
7/16/2008	open	<10-30	open	<10-20	open	<10	open	<10
7/17/2008	open	<10-30	open	<10-10	open	20	open	<10
7/18/2008	open	<10-20	open	<10-20	open	<10	open	<10
7/21/2008	open	10-40	open	<10-40	open	30	open	30
7/22/2008	open	<10-20	open	<10-20	open	10	open	10
7/23/2008	open	<10-20	open	<10-20	open	<10	open	10
7/24/2008	open	<10-30	open	<10-20	open	<10	open	<10
7/25/2008	open	<10-20	open	<10-20	open	20	open	<10
7/28/2008	open	10-20	open	<10-20	open	10	open	<10
7/29/2008	open	<10-20	open	<10-20	open	<10	open	<10
7/30/2008	open	<10-30	open	<10-30	open	31	open	<10
7/31/2008	open	10-30	open	<10-30	open	20	open	<10

# Numerical/Regulatory Agreement – *E. coli* (2008)

2008 <i>E. coli</i> , QPCR vs. Colilert-18 - Composite Samples, 1.0 m								
Date	QPCR Prediction (Status, CE)				Colilert-18 (Status, MPN)			
	North		Zoo		North		Zoo	
8/1/2008	open	10-50	open	<10-20	open	20	open	<10
8/4/2008	open	100-250	advisory	813	open	195	advisory	794
8/5/2008	open	10-30	open	30-70	open	20	open	63
8/6/2008	open	50-100	advisory	250-350	open	63	advisory	269
8/7/2008	open	30-60	open	<b>&lt;10-40</b>	open	52	open	<b>52</b>
8/8/2008	open	30-60	open	<10-20	open	63	open	20
8/11/2008	open	10-30	open	10-30	open	31	open	10
8/12/2008	open	20-50	open	<10-20	open	30	open	10
8/13/2008	open	30-70	open	23-30	open	63	open	31
8/14/2008	open	35-80	open	<10-20	open	84	open	10
8/15/2008	open	10-30	open	20-40	open	10	open	20
8/18/2008	open	20-50	open	<10-20	open	30	open	<10
8/19/2008	open	70-120	open	40-70	open	109	open	63
8/20/2008	open	10-30	open	40-70	open	20	open	63
8/21/2008	open	<10-20	open	20-30	open	<10	open	20
8/22/2008	open	<10-20	open	<b>&lt;10-30</b>	open	10	open	<b>52</b>
8/25/2008	<b>closed</b>	<b>&gt;1000</b>	<b>closed</b>	<b>&gt;1000</b>	<b>open</b>	<b>52</b>	<b>open</b>	<b>20</b>
8/26/2008	open	<10-20	open	<10-20	open	20	open	10
8/27/2008	open	<10-20	open	<10-20	open	10	open	<10
8/28/2008	open	20-40	open	30-60	open	31	open	63
8/29/2008	open	<b>&lt;10-20</b>	open	20-40	open	<b>31</b>	open	20
9/2/2008	open	<10-30	open	<10-20	open	<10	open	<10

# Correlation between Indicators - Cx

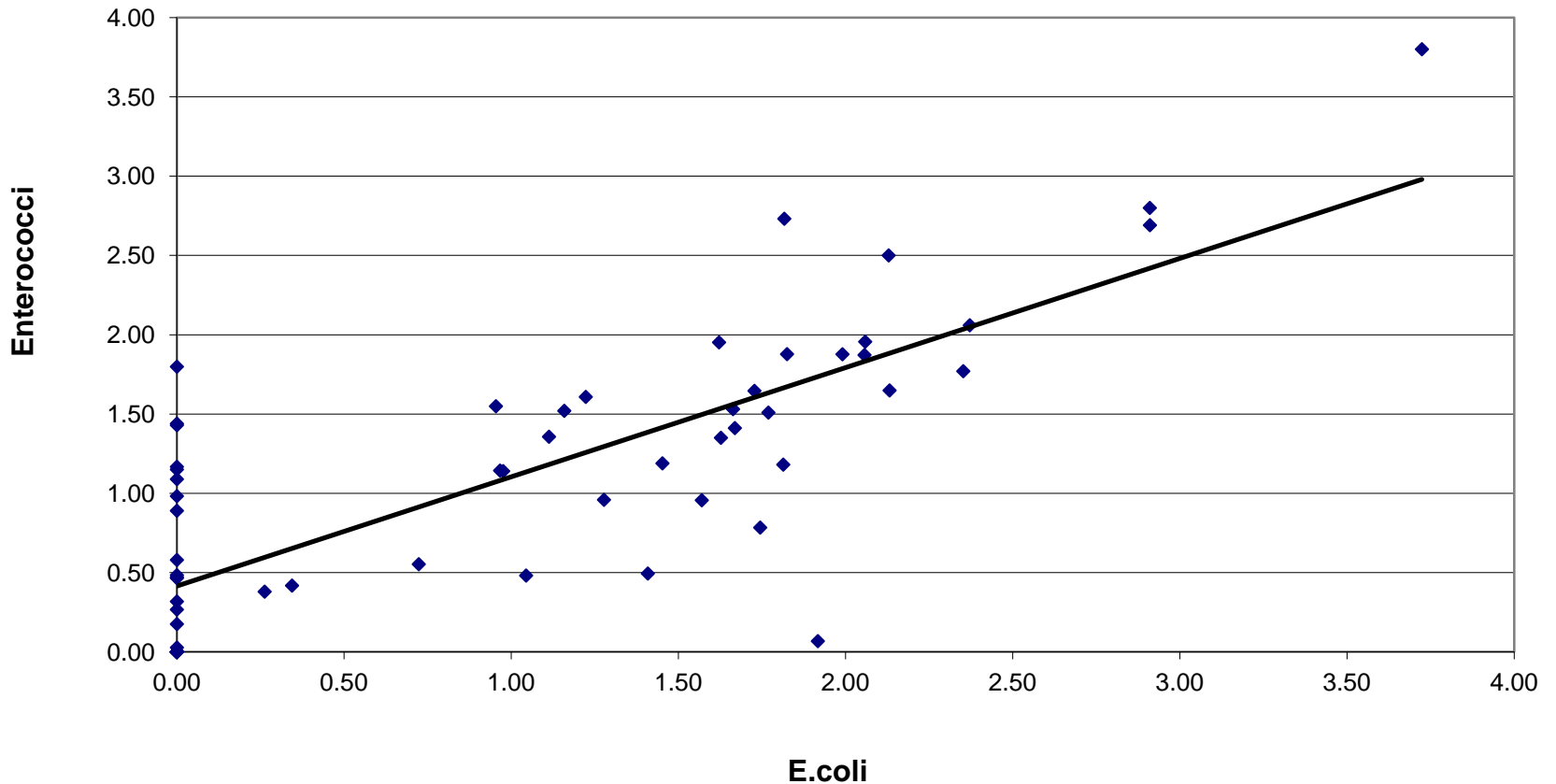
Enterococci (CFUs) and E.coli (MPN) Summer 08



# Correlation between Indicators - CCE

**E.coli (CCE) and Enterococci (CCE) Summer 08**  
Corrected CE Values

$$y = 0.6884x + 0.4158$$
$$R^2 = 0.618$$



# Results - 2008

- Enterococci – composite samples
  - 22 sampling days
  - Application of corrective factor when indicated
  - 98% agreement for regulatory action
  - 64% agreement for numerical value
- *E. coli* – composite samples
  - 46 sampling days
  - Application of corrective factor when indicated
  - 98% agreement for regulatory action
  - 87% agreement for numerical value

**2009 - 2010**

**Correlation w/o correction**

**Composite sampling**

## North Beach (2009) – *E. coli* vs. Enterococci

	<i>C-18</i>	<i>mEI</i>
C-18	1	
<b>mEI</b>	<b>0.910535</b>	<b>1</b>

## Zoo Beach (2009) – *E. coli* vs. Enterococci

	<i>C-18</i>	<i>mEI</i>
C-18	1	
<b>mEI</b>	<b>0.726051</b>	<b>1</b>

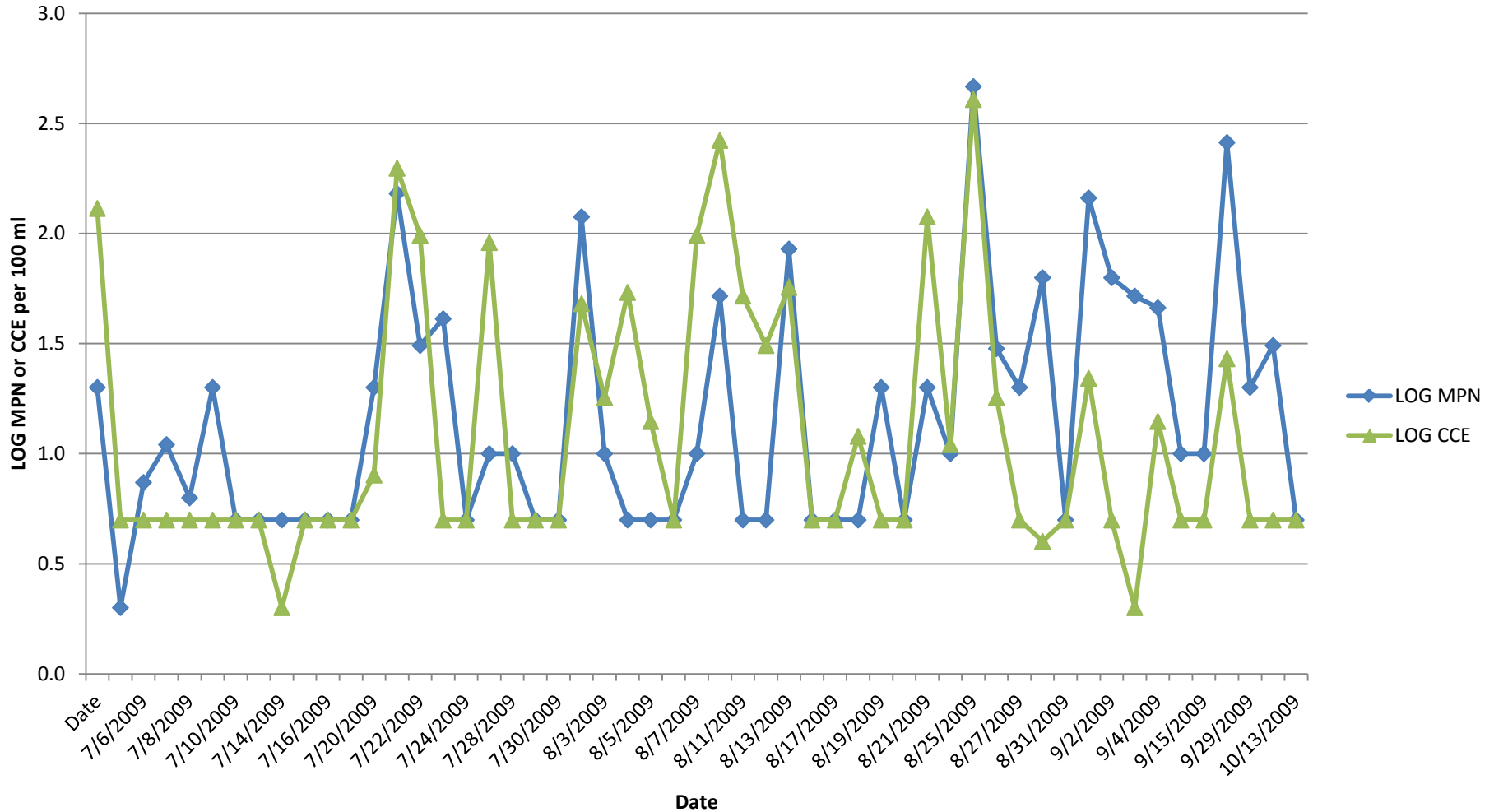
# Results – 2009

- North Beach
  - 53 sampling days
  - Numerical match, Colilert-18 vs. QPCR = 68%
  - Regulatory agreement = 96%\*
    - 1 Type I and 1 Type II error
- Zoo Beach
  - 52 sampling days
  - Numerical match, Colilert-18 vs. QPCR = 81%
  - Regulatory Agreement = 96%\*
    - 2 Type II errors

# North Beach - 2009

Date	Colilert-18	CCE- Uncorrected	CCE-UNC, MEAN	Date	Colilert-18	CCE- Uncorrected	CCE-UNC, MEAN
7/2/2009	<b>20</b>	119 - 140	<b>130</b>	8/6/2009	<10	11 - 17	14
7/6/2009	2.0	0	0	8/7/2009	<10	0	0
7/7/2009	7.4	0	0	8/10/2009	<b>10</b>	80 - 117	<b>98</b>
7/8/2009	11.0	0	0	8/11/2009	<b>52</b>	<b>220 - 310</b>	<b>265</b>
7/9/2009	6.3	0	0	8/12/2009	<10	0 - 52	26
7/10/2009	<b>20</b>	0	<b>0</b>	8/13/2009	<10	0 - 31	16
7/13/2009	<10	0	0	8/14/2009	<b>85</b>	0 - 57	<b>28</b>
7/14/2009	<10	0	0	8/17/2009	<10	0	0
7/15/2009	<10	0 - 2	1	8/18/2009	<10	0	0
7/16/2009	<10	0	0	8/19/2009	<10	8 - 17	12
7/17/2009	<10	0	0	8/20/2009	<b>20</b>	0	<b>0</b>
7/20/2009	<10	0	0	8/21/2009	<10	0	0
7/21/2009	20	0 - 8	4	8/24/2009	<b>20</b>	115 - 123	<b>119</b>
7/22/2009	152	93 - 304	198	8/25/2009	10	0 - 11	6
7/23/2009	31	32 - 164	98	8/26/2009	465	322 - 489	406
7/24/2009	<b>41</b>	0	<b>0</b>	8/27/2009	30	2 - 34	18
7/27/2009	<10	0	0	8/28/2009	20	0	0
7/28/2009	10	0 - 91	46	8/31/2009	<b>63</b>	2 - 7	<b>4</b>
7/29/2009	10	0	0	9/1/2009	<10	0	0
7/30/2009	<10	0	0	9/2/2009	<b>145</b>	16 - 28	<b>22</b>
7/31/2009	<10	0	0	9/3/2009	<b>63</b>	0	<b>0</b>
8/3/2009	<b>119</b>	44 - 52	<b>48</b>	9/4/2009	<b>52</b>	0 - 2	<b>1</b>
8/4/2009	10	7 - 29	18	9/8/2009	46	6 - 22	14
8/5/2009	<10	0 - 54	27	9/15/2009	10	0	0

## North Beach - E. coli, Colilert vs. QPCR (2009)

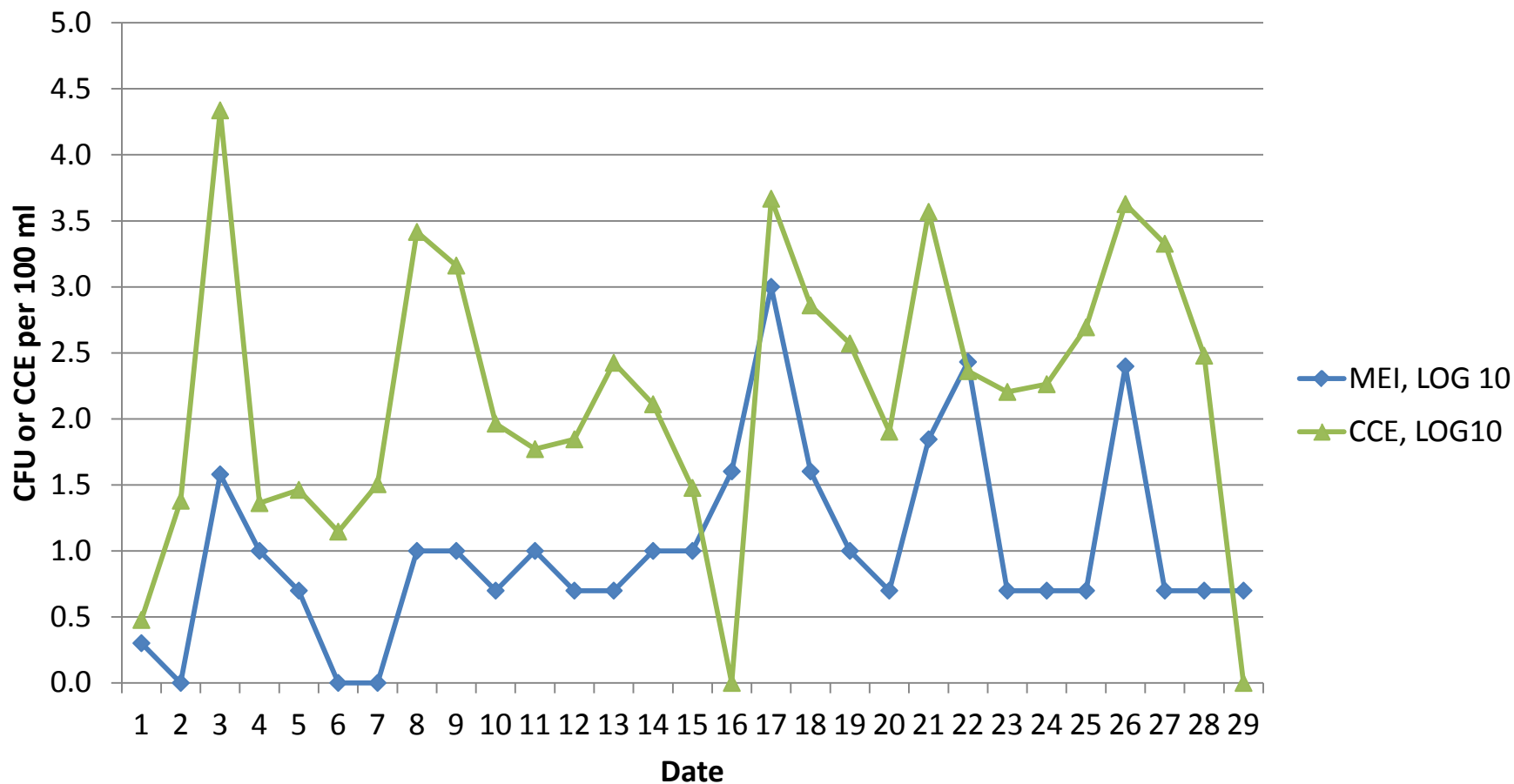


96% Regulatory Agreement

# Enterococci

NORTH	LOG cfu	CCE	LOG/CCE
LOG cfu	1		
CCE	0.362849411	1	
<b>LOG/CCE</b>	<b>0.50710848</b>	0.587393	<b>1</b>

## North Beach - Enterococci, mEI Agar vs. QPCR (2009)



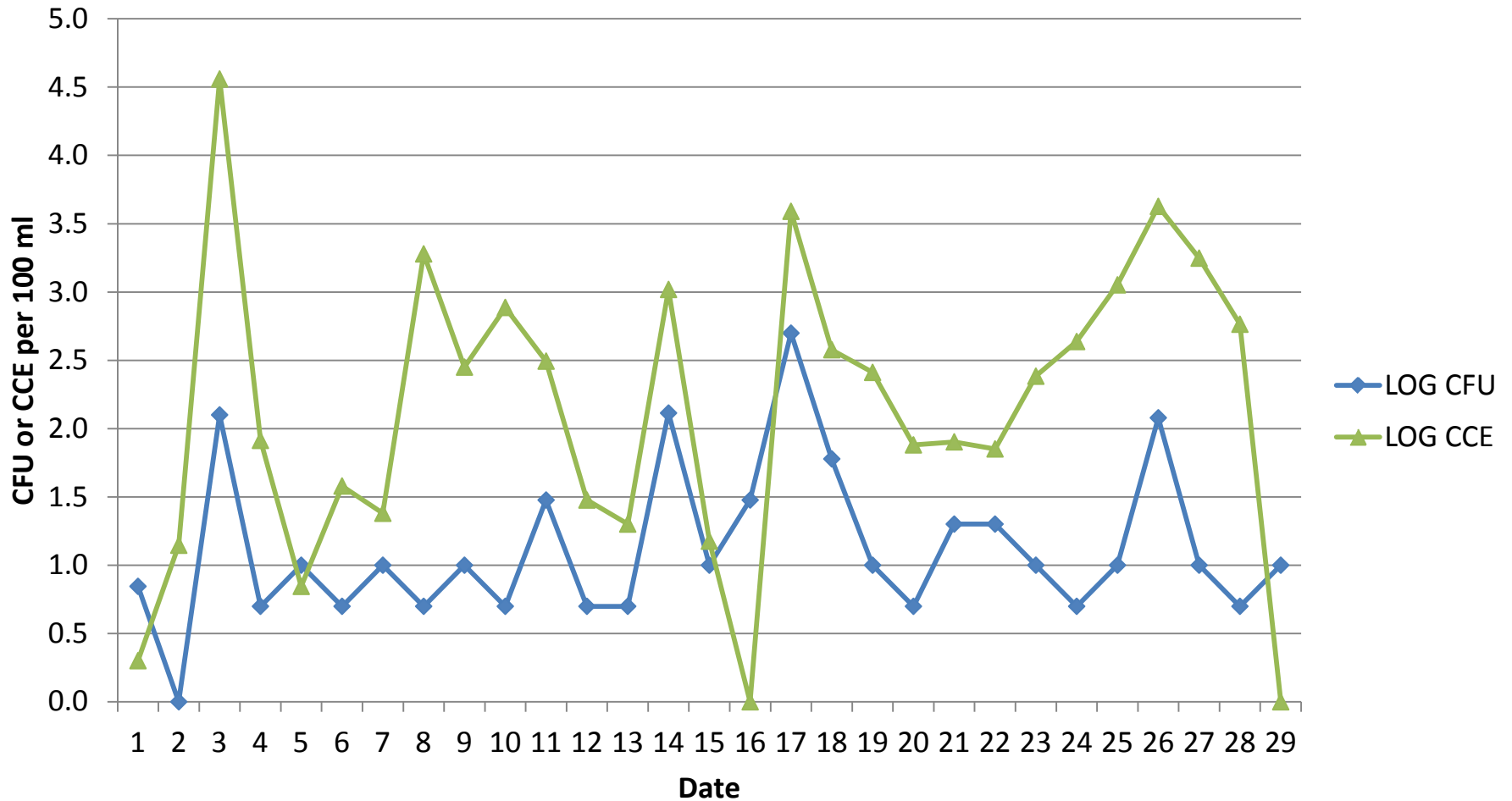
# Zoo Beach - 2009

Date	Colilert-18	CCE- Uncorrected	CCE-UNC, MEAN	Date	Colilert-18	CCE- Uncorrected	CCE-UNC, MEAN
7/2/2009	<b>689</b>	309 - 449	<b>379</b>	8/6/2009	<10	0	0
7/6/2009	1.0	0	0	8/7/2009	<b>85</b>	0	<b>0</b>
7/7/2009	7.4	0	0	8/10/2009	<10	0 - 6	3
7/8/2009	8.5	0	0	8/11/2009	<b>10</b>	111 - 244	<b>178</b>
7/9/2009	1.0	0	0	8/12/2009	<10	0	0
7/10/2009	74	39	39	8/13/2009	<b>&lt;10</b>	64 - 144	<b>104</b>
7/13/2009	30	54 - 75	64	8/14/2009	269	401	401
7/14/2009	<10	0 - 23	12	8/17/2009	20	0 - 29	14
7/15/2009	<b>63</b>	0 - 11	<b>6</b>	8/18/2009	<10	0 - 4	2
7/16/2009	<10	0	0	8/19/2009	<10	0	0
7/17/2009	<10	0	0	8/20/2009	<b>63</b>	6	<b>6</b>
7/20/2009	<10	0	0	8/21/2009	<b>20</b>	0	<b>0</b>
7/21/2009	<b>132</b>	64 - 71	<b>68</b>	8/24/2009	<10	0	0
7/22/2009	85	29 - 86	58	8/25/2009	10	0 - 3	2
7/23/2009	10	12 - 106	59	8/26/2009	313	256 - 271	264
7/24/2009	<10	0	0	8/27/2009	20	0	0
7/27/2009	<10	0	0	8/28/2009	<b>30</b>	0	<b>0</b>
7/28/2009	<10	8 - 40	24	8/31/2009	<b>20</b>	0 - 1	<b>1</b>
7/29/2009	<10	0	0	9/1/2009	<10	0	0
7/30/2009	<10	0 - 2	1	9/2/2009	<b>31</b>	0	<b>0</b>
7/31/2009	<10	0	0	9/3/2009	<10	0	0
8/3/2009	<b>404</b>	160 - 302	<b>231</b>	9/4/2009	20	0 - 1	1
8/4/2009	<10	0 - 14	7	9/8/2009	6	0 - 5	3
8/5/2009	<10	0	0	9/15/2009	10	0	0

# *Enterococci*

ZOO	LOG,CFU	CCE	LOG/CCE
LOG/CFU	1		
CCE	0.419861	1	
<b>LOG/CCE</b>	<b>0.462341</b>	0.517691	1

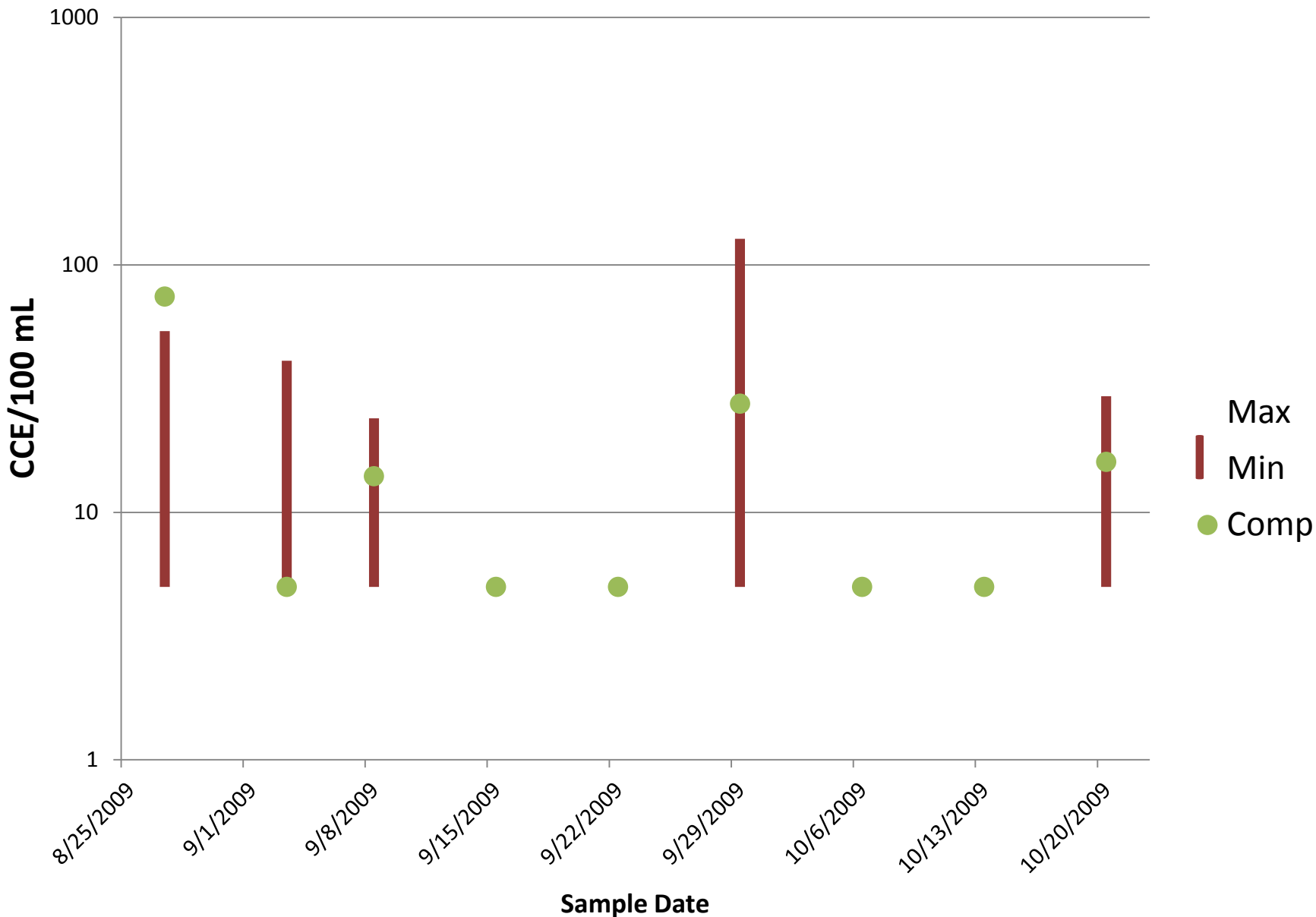
## Zoo Beach - Enterococci, mEI Agar vs. QPCR (2009)



# Composite Sampling

- August 2009 – June 2010
  - No mean difference in *E. coli* concentration across beach transects, i.e. valid to composite samples
  - 40 sampling events across two beaches
  - Composite samples fell within the range of the individual samples 92% of the time
  - The use of composite sampling would have made no difference with respect to regulatory action
- Compositing samples represents a significant cost savings

# North Beach *E. coli* Composite vs. Individual Site Analysis, QPCR



# Results – 2010

- *E. coli*
- North Beach
  - 87 sampling days
  - Numerical match, Colilert-18 vs. QPCR = 66%
  - Regulatory Agreement = 98%\*
    - 1 Type I and 1 Type II error
- Zoo Beach
  - 88 sampling days
  - Numerical match, Colilert-18 vs. QPCR = 73%
  - Regulatory Agreement = 86%\*
    - 12 Type I errors
    - \*Cluster of 8 errors likely due to analyst error

# 2010 QPCR Implementation Pilot Study

- July 23 – September 7, 2010
- Parallel to similar effort in Orange County, CA
- Daily management decisions made same-day using QPCR
- Confirmed the following morning by culture
- Non-consensus or inhibition defaulted to previous day's *E. coli* results as determined by the culture-based method
- Same management decisions would have been made for the duration of the pilot study, regardless of analytical method

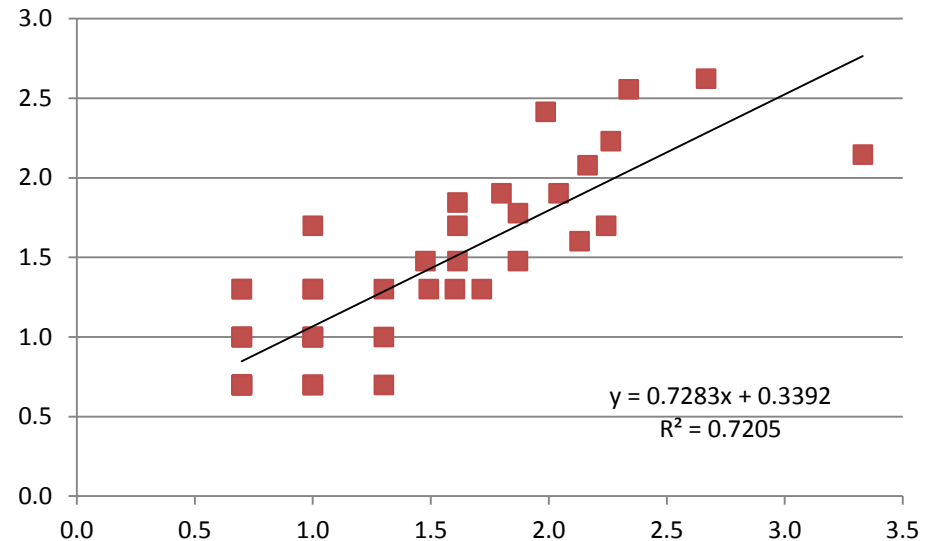
# 2011 Final Pre-Implementation Study

- Establish relationship to NEEAR health effects
  - Enterococci and *E. coli* by culture
- Establish relationship between qPCR and culture
  - Enterococci by culture and qPCR
  - *E. coli* by culture and qPCR
- Determine if sampling time was reflective of water quality
- Examine regulatory action agreement
  - Choice of indicator
  - Do results make sense in light of sanitary survey data
- Look at qPCR vs. Virtual Beach

# Correlation: culture to culture

There is not perfect agreement, even between two culture-based methods

C-18 vs. 1603, North Beach - AM (2011)



Anova: Single Factor

## SUMMARY

Groups	Count	Sum	Average	Variance
Colilert-18	46	4388	95.39130435	102011.8879
Method 1603	46	2350	51.08695652	7865.458937

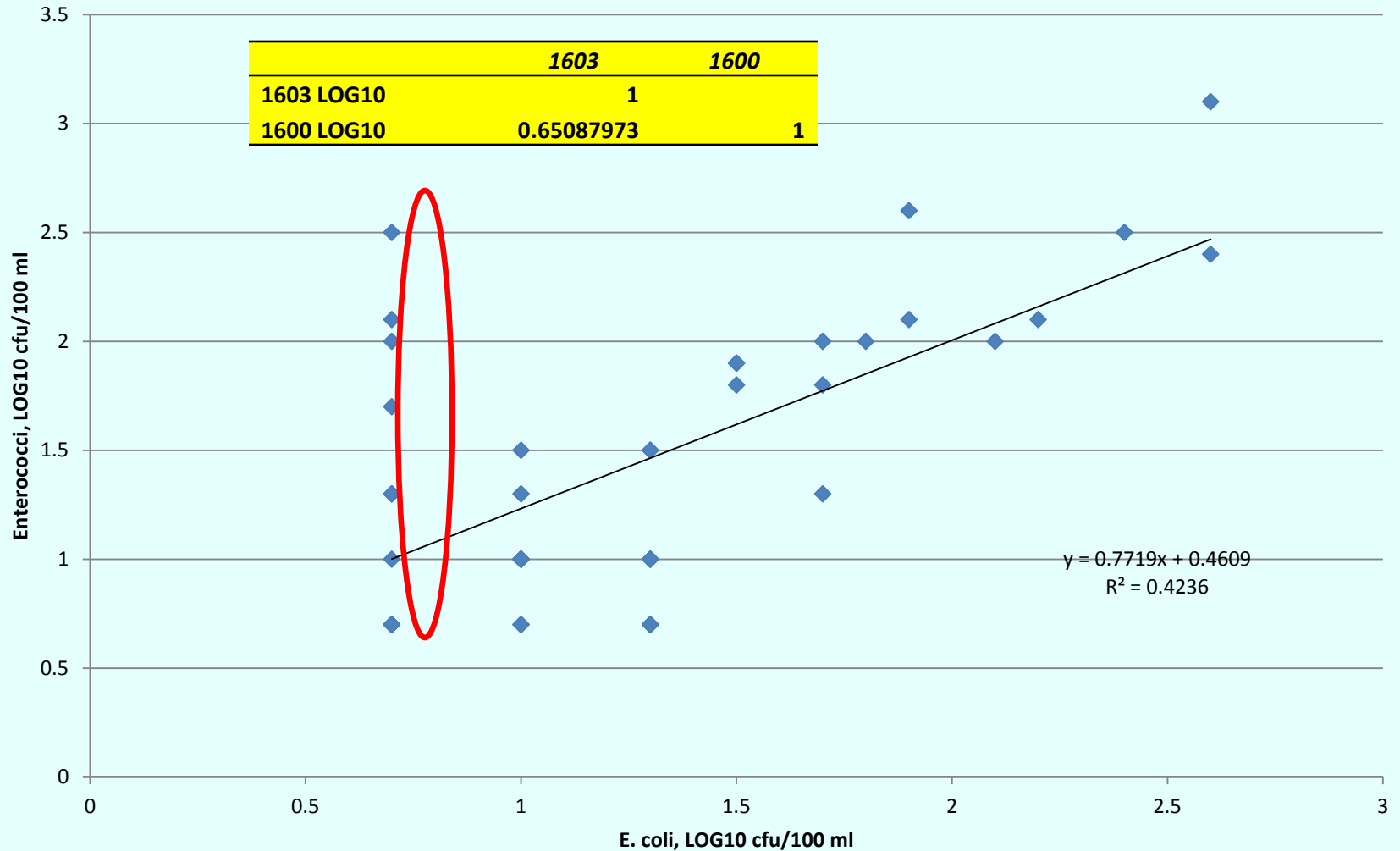
## ANOVA

Source of Variation	SS	df	MS	F	P-value	F crit
Between Groups	45146.13043	1	45146.13043	0.821755015	0.367088177	3.946875558
Within Groups	4944480.609	90	54938.67343			
Total	4989626.739	91				

**METHOD 1603**  
**vs.**  
**Colilert-18**

# *E. coli* (Method 1603) and enterococci (Method 1600) were correlated when using culture-based enumeration methods

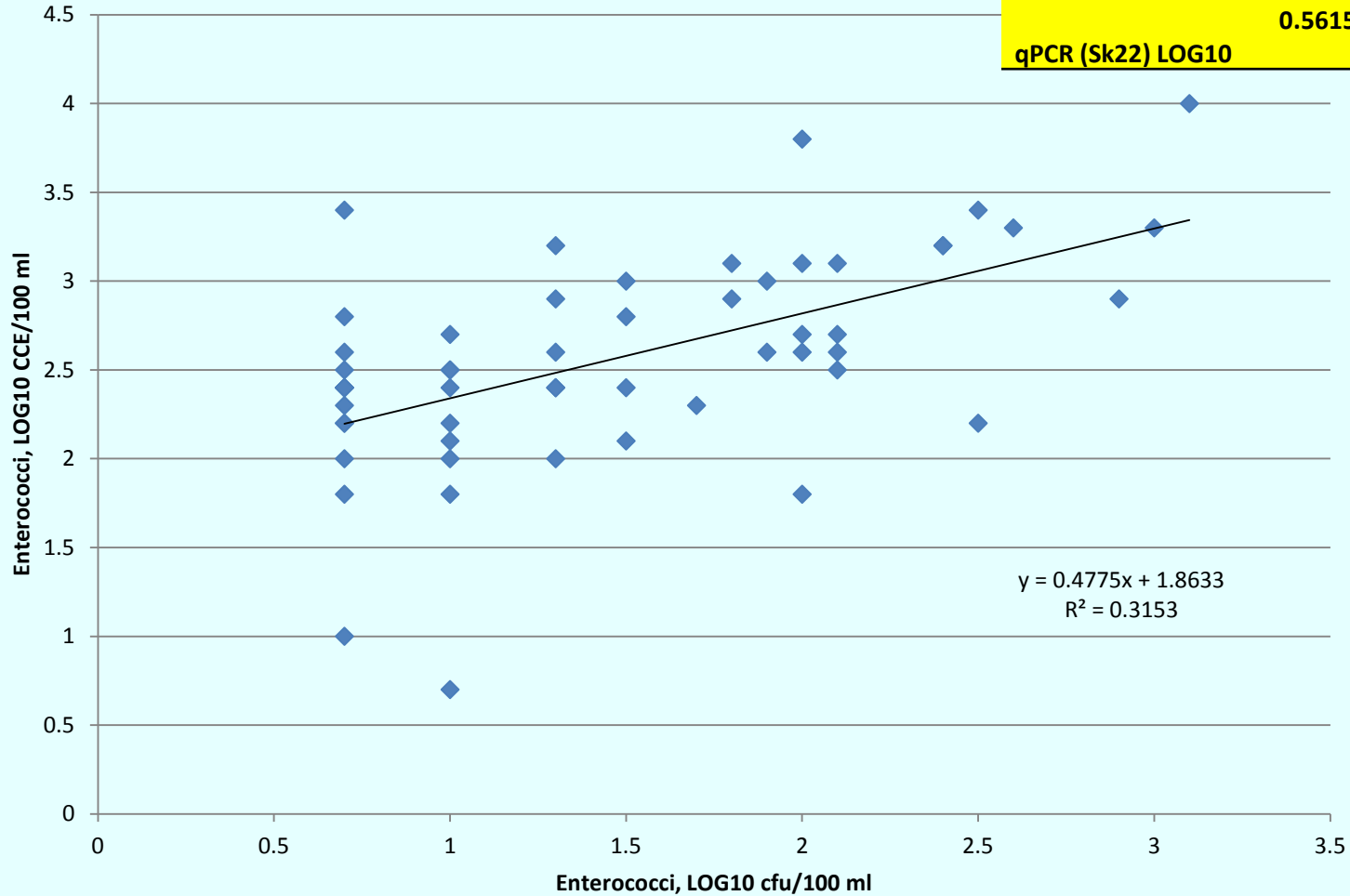
## Log10 Method 1603 vs. Method 1600 (2011)



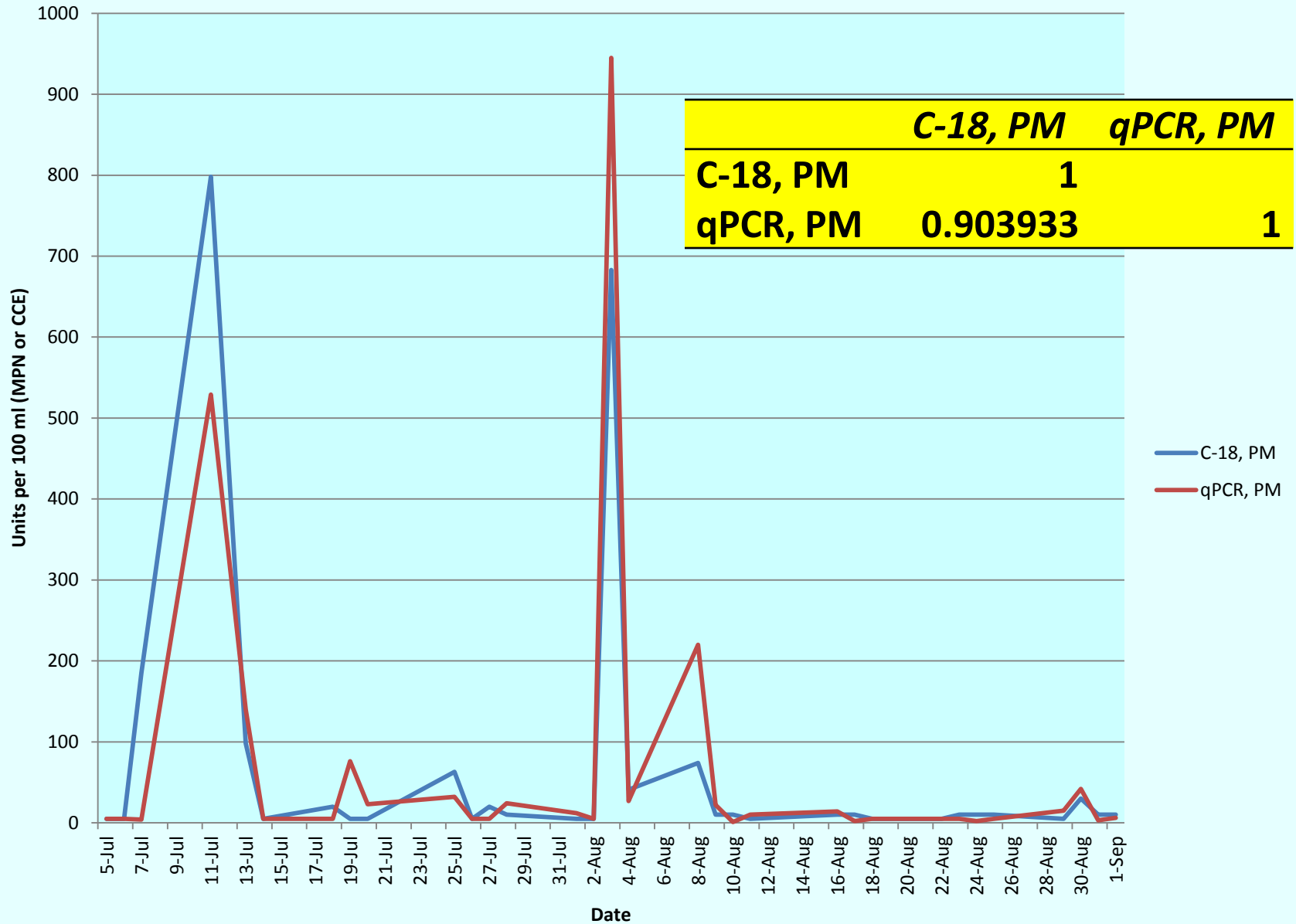
# Enterococci, as enumerated by qPCR (CCE/100 ml), was positively correlated with Enterolert.

Method 1600 vs. qPCR LOG10

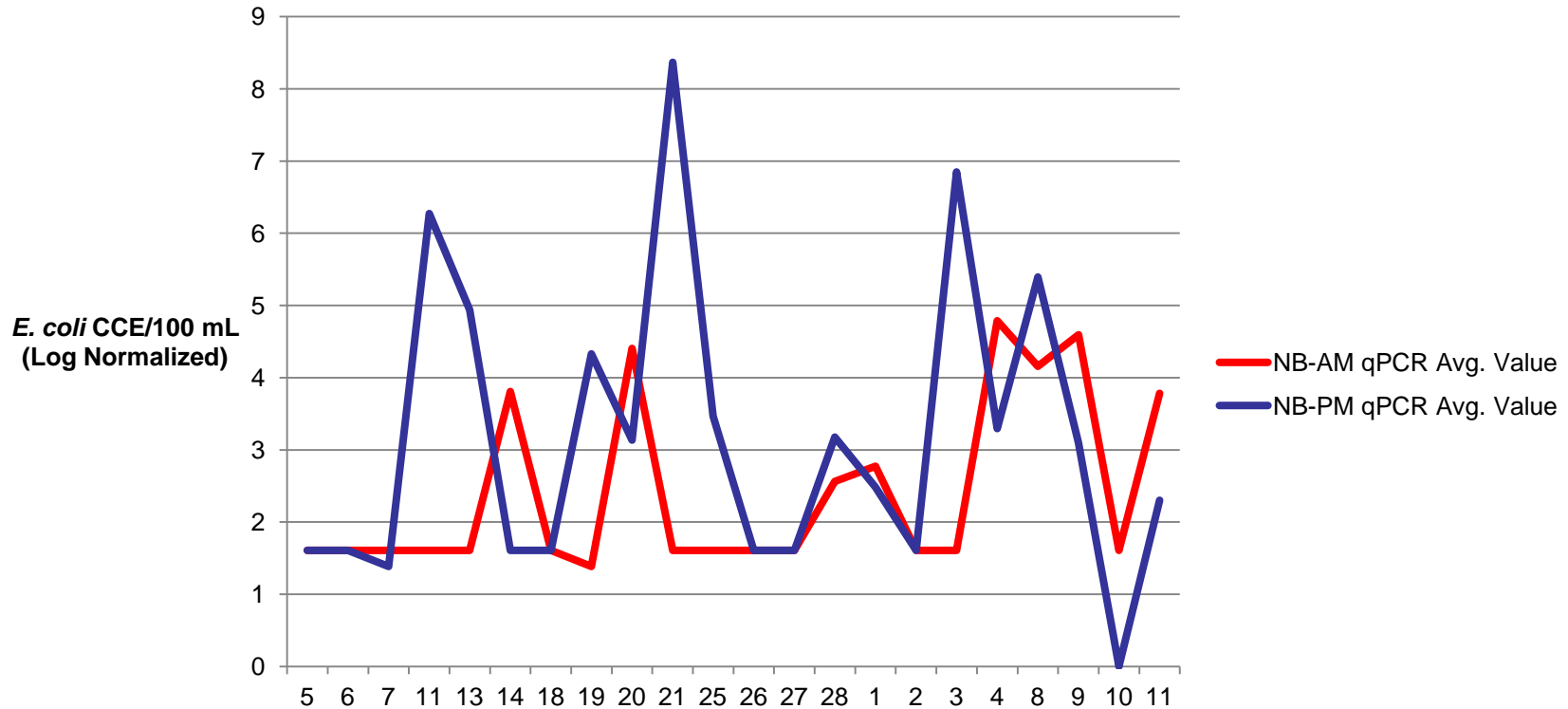
	Method 1600	qPCR
Method 1600 LOG10	1	
	0.5615192	
qPCR (Sk22) LOG10	43	1



## *E. coli*, Colilert-18 vs. qPCR (2011)



## ***E. coli* CCE/100 mL vs. Date of Sample (2011)**



**TABLE 1 - ANOVA: Single Factor, qPCR**

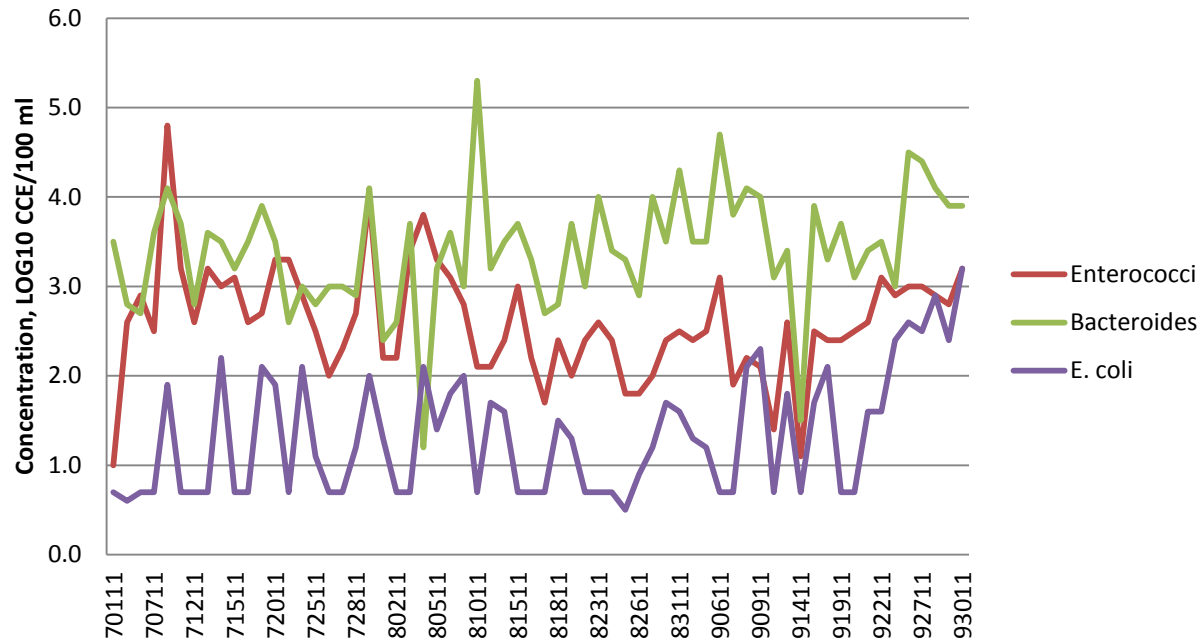
**SUMMARY**

Groups	Count	Sum	Average	Variance
qPCR AM	22	53.18559135	2.41752688	1.468841003
qPCR PM	22	69.76056173	3.170934624	4.343360239

**ANOVA**

Source of Variation	SS	df	MS	F	P-value	F crit
Between Groups	6.243855525	1	6.243855525	2.148533839	0.150153276	4.072653663
Within Groups	122.0562261	42	2.906100621			
Total	128.3000816	43				

## Enterococci, Bacteroides, and *E. coli* (2011)



The use of enterococci would have resulted in significantly more exceedances.

For culture-based assays, the use of enterococci would have resulted in 7 (IDEXX Colilert-18 vs. Enterolert) to 13 (USEPA Method 1603 vs. 1600) more advisories based on proposed SVT values of 235 and 61 for *E. coli* and enterococci respectively.

For qPCR, the use of enterococci would have resulted in 10 additional exceedances based on a an SVT of 235 CCE/100 ml for *E. coli* (use of same criteria but alternative analytic method as per p. 55) and 1000 CCE/100 ml for enterococci.

# Remember qPCR is *not* Measuring the Same Thing as a Culture...

- qPCR differs from traditional culture-based assays in that it measures all DNA:
  - live cells
  - dead cells
  - non-culturable cells
  - free DNA
- Culture assays only measure cells possessing the ability to grow on the selective media you are using
- Water body characteristics and environmental conditions may effect
  - Inhibition
  - Culture/qPCR relationship
  - Ability to use qPCR as an analytical method

# Predictive Models

- Environmental data collected as part of a Beach Sanitary Survey can be used to begin constructing a predictive model
- Virtual Beach (US EPA model)
  - <http://www.epa.gov/ceampubl/swater/vb2/vb2-2-dl.html>
  - Allows correlations between parameters and water quality (i.e. wave height, wind direction, rainfall, etc.)
  - Can be combined with hydrodynamic impact models to look at influence of land use on fecal loading (L-THIA)
  - Data limitations prevent full use, i.e. you will likely need multiple years of data
- Other models available:
  - SwimCast (Illinois), Nowcast (Ohio), Project SAFE (Indiana), Rainfall-based alerts (Wisconsin, others)

# VB helps turn data like these...

Microsoft Excel - HBSe1 - rev.xls

File Edit View Insert Format Tools Data Window Help

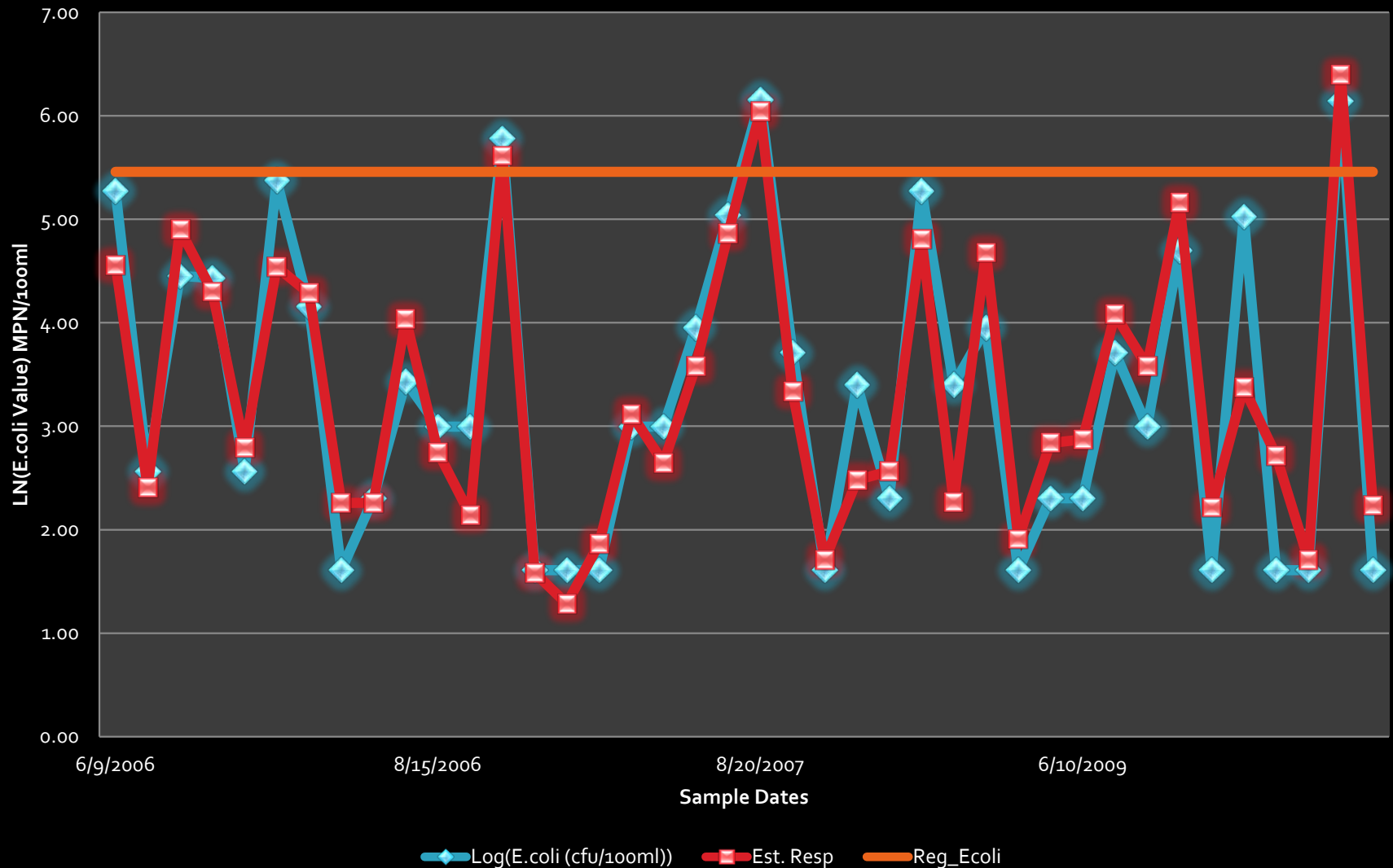
Type a question for help

AJ103

	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	Y	Z	AA	AB	AC	AD	AE	AF	AG	AH	AI	AJ	AK			
1	Date	Time	E-coll	24T	24DP	24cld	24vws	24dir	24sp	24pot	Ufcst	Vfcst	U20rot	V20rot	turbidit	temp	wav	24hrR	48hrR	SolarD	Solar8	Solar9	dirN=C	wind	U	V	9amT	9aDP	clot	samp1	Julian	Date	8aDP	mb9aCLE	mb9aBoy	mb8aBoy	del			
2	9/3/2006						1.75																																	
3	9/2/2006				62	55	87	7.00	45	23	61	-16.26	-16.26	-20.85	-9.72																									
4	9/1/2006	8:36			66	56	80	3.25	90	13	22	-13.00	0.00	-12.22	4.45	104.3	73	3	0.01	0.01	278	96	233	90	12.0	-12.0	0.0													
5	8/31/2006	9:02	520	66	57	80	5.25	68	17	40	-15.76	-6.37	-16.99	-0.59	83.0	74	3	0.00	0.02	439	110	308	90	13.0	-13.0	0.0	64	53	4	122	243	8/31/2006	49	1019.6	1018.4	1017.6				
6	8/30/2006	8:52	1667	67	62	63	4.75	45	17	20	-12.02	-12.02	-15.41	-7.18	162.8	74	2	0.02	1.27	380	133	355	90	12.0	-12.0	0.0	68	60	4	112	242	8/30/2006	66	1015.7	1014.5	1014.2				
7	8/29/2006	9:52	3850	69	64	86	5.50	338	9	46	3.37	-8.34	0.31	-8.99	258.0	76	4	1.25	1.53	172	16	48	360	13.0	0.0	-13.0	68	66	5	172	241	8/29/2006	66	1008.5	1007.4	1008.1				
8	8/28/2006	8:54	86	70	65	79	0.25	22	5	30	-1.87	-4.64	-3.35	-3.72	32.2	76	1	0.28	0.33	124	16	89	0	0.0	0.0	0.0	69	65	5	114	240	8/28/2006	68	1015.4	1015.0	1014.5				
9	8/27/2006	8:49	349	73	67	89	2.25	202	8	79	3.00	7.42	5.35	5.95	49.0	76	1	0.05	0.05	166	23	41	0	0.0	0.0	0.0	71	68	5	109	239	8/27/2006	68	1013.5	1012.7	1012.6				
10	8/26/2006	8:31	273	70	64	36	1.00	180	5	17	0.00	5.00	1.71	4.70	44.4	76	2	0.00	0.02	331	151	314	180	3.0	0.0	3.0	72	65	4	91	238	8/26/2006	66	1017.0	1015.9	1015.2				
11	8/25/2006	8:30	21	69	62	67	0.75	158	13	29	-4.87	12.05	-0.45	12.99	3.5	76	1	0.02	0.02	334	127	316	180	5.0	0.0	5.0	72	64	4	90	237	8/25/2006	58	1016.1	1014.1	1014.2				
12	8/24/2006	8:53	45	67	57	74	0.75	158	8	44	-3.00	7.42	-0.28	8.00	5.9	76	1	0	0	243	66	40	225	3.0	2.1	2.1	65	57	5	113	236	8/24/2006	60	1018.6	1014.5	1014.3				
13	8/23/2006	9:04	114	65	59	40	0.75	22	7	14	-2.62	-6.49	-4.68	-5.20	6.2	77	1	0	0	454	182	346	135	3.0	-2.1	2.1	69	60	2	124	235	8/23/2006	60	1018.6	1017.6	1017.4				
14	8/22/2006	9:17	11	66	57	40	0.75	225	7	16	4.95	4.95	6.34	2.96	9.0	77	1	0	0	557	188	372	225	6.0	4.2	4.2	65	56	2	137	234	8/22/2006	56	1021.6	1020.6	1020.0				
15	8/21/2006	8:46	93	65	57	18	0.75	45	5	0	-3.54	-3.54	-4.53	-2.11	32.9	74	1	0	0.31	459	184	351	0	0.0	0.0	0.0	62	57	1	106	233	8/21/2006	57	1021.9	1020.8	1020.8				
16	8/20/2006	8:36	382	71	65	67	1.25	360	9	14	0.00	-9.00	-3.08	-8.46	29.8	75	2	0.31	0.32	303	52	125	315	9.0	6.4	-6.4	69	62	5	96	232	8/20/2006	62	1015.2	1014.4	1013.7				
17	8/19/2006	8:25	27	69	64	81	1.75	180	8	71	0.00	8.00	2.74	7.52	3.5	76	1	0.01	0.01	110	16	93	225	9.0	6.4	6.4	72	70	5	85	231	8/19/2006	70	1014.9	1012.9	1013.6				
18	8/18/2006	8:43	50	71	62	53	0.75	180	9	10	0.00	9.00	3.08	8.46	6.8	78	1	0	0	174	64	113	180	7.0	0.0	7.0	73	64	5	103	230	8/18/2006	63	1019.8	1018.4	1018.4				
19	8/17/2006	8:54	32	67	63	11	0.75	135	5	5	-3.54	3.54	-2.11	4.53	5.3	79	1	0	0	537	199	404	135	6.0	-4.2	4.2	71	55	3	114	229	8/17/2006	55	1023.6	1022.5	1022.8				
20	8/16/2006	8:57	3	66	58	9	1.25	45	5	2	-3.54	-3.54	-4.53	-2.11	6.9	77	1	0	0.12	595	210	391	0	0.0	0.0	0.0	69	59	1	117	228	8/16/2006	58	1022.8	1021.6	1021.3				
21	8/15/2006	8:22	309	68	62	24	2.00	248	11	4	10.20	4.12	10.99	0.38	28.8	79	2	0.12	0.12	586	214	393	225	7.0	4.9	4.9	66	60	4	82	227	8/15/2006	58	1015.6	1014.2	1013.6				
22	8/14/2006	8:48	13	71	63	41	2.75	202	14	14	5.24	12.98	9.37	10.40	6.8	77	1	0	0	360	175	293	225	12.0	8.5	8.5	71	58	1	108	226	8/14/2006	54	1014.1	1012.4	1012.7				
23	8/13/2006	8:24	126	63	56	9	1.58	5	2	-1.87	4.64	-0.17	5.00	11.6	79	1	0	0	606	227	421	0	0.0	0.0	0.0	63	53	2	84	225	8/13/2006	51	1021.1	1020.0	1019.8					
24	8/12/2006	7:52	32	63	49	14	1.75	112	8	1	-7.42	3.00	-5.95	5.35	33.8	77	2	0	0	593	230	424																		
25	8/11/2006	8:22	387	68	58	41	4.25	68	15	3	-13.91	-5.62	-14.99	-0.52	107.9	78	2	0	0	611	214	416	90	13.0	-13.0	0.0	72	52	3	82	223	8/11/2006	54	1016.0	1015.2	1014.4				
26	8/10/2006	9:04	28	68	62	36	2.02	5	14	1.87	4.64	3.35	3.72	7.0	79	1	0	0	517	187	267	0	0.0	0.0	0.0	71	61	5	124	222	8/10/2006	60	1014.3	1013.4	1013.4					
27	8/9/2006	8:49	57	65	61	12	0.25	135	7	12	-4.95	4.95	-2.96	6.34	26.5	80	1	0	0	615	237	425	135	6.0	-4.2	4.2	67	51	3	109	221	8/9/2006	54	1021.7	1020.6	1020.6				
28	8/8/2006	8:50	284	70	60	19	4.5	9	19	-6.36	-6.36	-8.16	-3.80	35.6	79	2	0	0.05	536	186	207	360	13.0	0.0	-13.0	73	58	3	110	220	8/8/2006	59	1022.2	1021.6	1021.5					
29	8/7/2006	8:58	84	77	69	75	2.25	248	11	31	10.20	4.12	10.99	0.38	4.5	80	1	0.05	0.05	434	161	331	225	8.0	5.7	5.7	77	70	4	118	219	8/7/2006	69	1018.4	1017.2	1016.7				
30	8/6/2006	7:55	56	73	62	33	0.75	158	8	13	-3.00	7.42	-0.28	8.00	13.3	78	1	0	0	527	234	406	180	9.0	0.0	9.0	73	59	3	55	218	8/6/2006	59	1019.5	1018.4	1018.3				
31	8/5/2006	8:40	135	70	62	14	0.75	112	6	5	-5.56	2.25	-4.46	4.01	40.1	79	2	0	0.09	591	235	419	90	7.0	-7.0	0.0	74	56	2	100	217	8/5/2006	59	1022.0	1021	1020.7				
32	8/4/2006	8:20	326	74	64	47	0.25	360	8	8	0.00	-8.00	-2.74	-7.52	40.6	78	3	0.09	0.09	613	241	417	360	12.0	0.0	-12.0	73	64	3	80	216	8/4/2006	61	1017.5	1016.8	1016				
33	8/3/2006	9:07	463	79	76	56	0.75	270	10	48	10.00	0.00	9.40	-3.42	7.8	80	1	0	0	288	221	417	225	9.0	6.4	6.4	81	72	1	127	215	8/3/2006	72	1011.2	1010.4	1009.4				
34	8/2/2006	9:18	17	83	73	32	1.25	225	11	14	7.78	7.78	9.97	4.65	3.2	81	1	0	0.01	553	224	394	225	12.0	8.5	8.5	82	73	1	138	214	8/2/2006	72	1014.5	1012.8	1012.9				
35	8/1/2006	9:04	21	82	73	19	0.25	225	8	5	5.66	5.66	7.25	3.38	2.0	81	1	0.01	0.47	577	211	381	225	10.0	7.1	7.1	82	74	2	124	213	8/1/2006	73	1015.7	1014.2	1014.1				
36	7/31/2006	9:04	65	76	72	42	0.75	225	9	14	6.36	6.36	8.16	3.80	2.7	80	1	0.46	0.46	558	150	360	225	7.0	4.9	4.9	78	72	5	124	212	7/31/2006	71	1014.0		1011.2				
37	7/30/2006	8:43	38	74	73	49	0.75	225	8	29	5.66	5.66	7.25	3.38	4.4	80	1	0	0	475	152	336	270	5.0	5.0	5.0	79	72	3	103	211	7/30/2006	71	1014.0						
38	7/29/2006	8:26	69	77	68	39	0.25	225	5	14	3.54	3.54	4.53	2.11	2.5	79	1	0	0.12	548	203	394	225	7.0	4.9	4.9	75	70	2	86	210	7/29/2006	70	1015.7	1015	1014.3				
39	7/28/2006	8:26	567	76	70	74	0.75	225	6	37	4.24	4.24	5.44	2.54	9.9	76	1	1.12	1.49	511	108	128	270	10.0	10.0	0.0	70	67	5	86	209	7/28/2006	67	1015.0	1013.4	1012.2				
40	7/27/2006	9:03	121	75	68	69	1.25	180	14																															

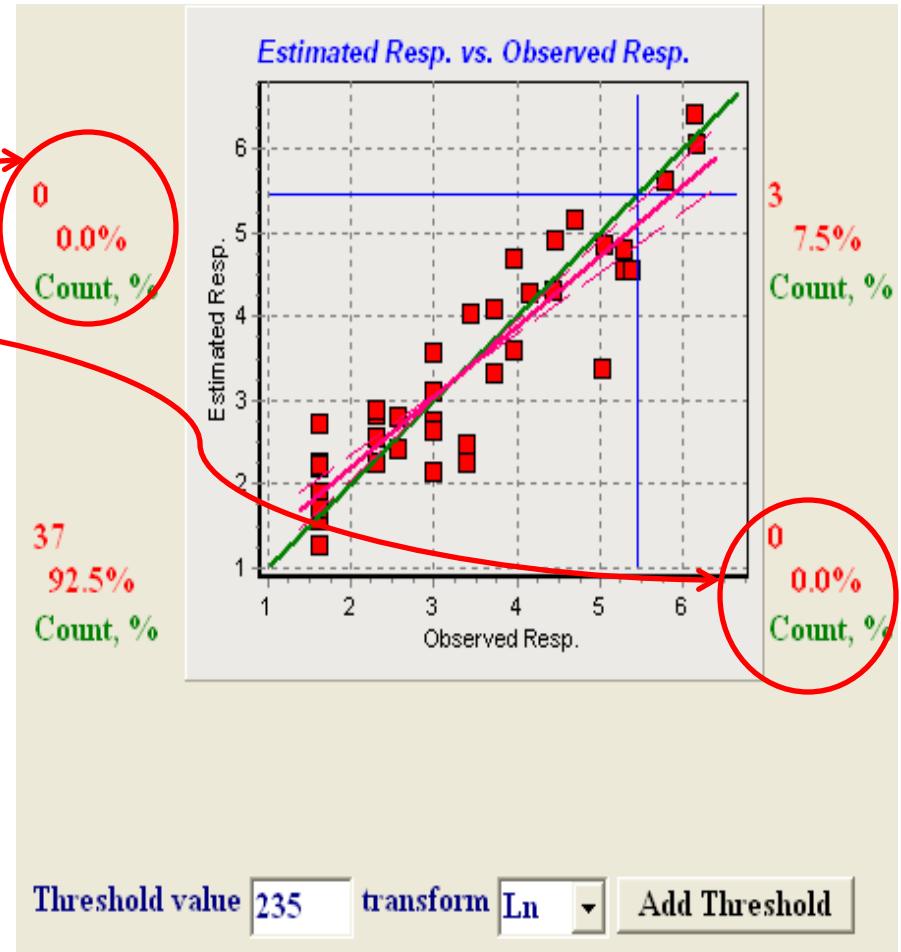
# ...into estimated *E. coli* counts

(1) Actual v. Predicted ( 2006-2009)



# North Beach Model- Training

- $R^2 = 0.793$ ,  $n = 40$
- Database covers years from 2006-2009
- No observed false positives or false negatives
- Variables include:
  - Algae Description
    - Floating or Submerged
  - # Gulls
  - Wave Height
  - Wind Speed
  - 24 hr rainfall
  - NE Winds
  - Interactive variable



# 2011 VB Model Pilot Study

## VB Predictions vs. Colilert-18

Correct, 42/46	91%
Incorrect, 4/46	9%
Type I, 2/46	4%
Type II, 2/46	4%

## VB Predictions vs. qPCR CCE

Correct, 43/44	98%
Incorrect, 1/44	2%
Type I, 1/44	2%
Type II, 0/46	0%

The predictive model constructed using Virtual Beach accurately estimated *E. coli* concentrations enumerated by culture-based (Colilert-18, 91%) and qPCR assays (98%).

# Limitations

- Success of model is limited to the availability of quality data and the overall history of the beach
- Difficulty lies with consistency and number of parameters monitored
- Sanitary Survey data greatly improves the chances of a successful and quality model being produced
  - Greater quantity of data for each monitoring day
  - Much more detail associated with each observed parameter

# Implementation - 2012

- Regulatory monitoring 5 days/week
  - Received approval as an ATP indicator/method combination in May 2012
  - 2 beaches
  - *E. coli* by qPCR (BioGx Smartbeads™: *E. coli* and Sketa)
  - Compare to culture 4 days/week
  - Compare to model 5 days/week
  - Compare *E. coli*/enterococci by culture (qPCR to follow)
    - Archived filters (Fall 2012)

# Murphy's Law...sort of

<b>SITE</b>	<b>Numerical Correlation (r)</b>	<b>Regulatory Agreement</b>	<b># Advisories E. coli - Cx</b>	<b>#Advisories Alternative</b>
<b>North Beach</b>				
<b>Colilert vs. Enterolert</b>	<b>0.94</b>	<b>88%</b>	<b>5</b>	<b>9</b>
<b>Colilert vs. qPCR</b>	<b>0.59</b>	<b>89%</b>	<b>6</b>	<b>12</b>
<b>qPCR vs. Virtual Beach</b>	<b>0.19</b>	<b>78%</b>		
<b>Zoo Beach</b>				
<b>Colilert vs. Enterolert</b>	<b>0.97</b>	<b>96%</b>	<b>3</b>	<b>5</b>
<b>Colilert vs. qPCR</b>	<b>0.46</b>	<b>98%</b>	<b>6</b>	<b>7</b>
<b>qPCR vs. Virtual Beach</b>	<b>0.49</b>	<b>91%</b>		

# Data Handling Reality Check

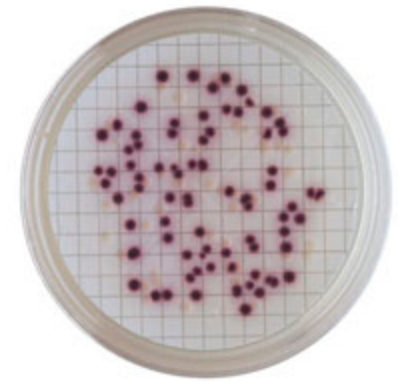
- Replicate analyses do not always agree
  - Open, advisory, closed, coin toss (?)
- Culture does not always agree with qPCR
- Results do not always make sense based on environmental conditions
- Inhibition
- False negatives w/o inhibition (algae blooms?)
- Filter blanks or NTC are contaminated
- Contamination as a result of visitors/observers not wearing PPE

# **Recommended Course of Action**

**Based on our experiences with  
implementation**

# Remember QPCR is *not* Measuring the Same Thing as a Culture...

- QPCR differs from traditional culture-based assays in that it measures all DNA:
  - live cells
  - dead cells
  - non-culturable cells
  - free DNA
- Culture assays only measure cells possessing the ability to grow on the selective media you are using



# Feasibility Studies

- Ability of the method to accurately characterize water quality
  - Will you use the same indicator as your culture-based assay?
  - Will the assay work with your samples?
    - Inhibition, underestimation
    - Unexplained false negatives w/o inhibition
- Turn-Around-Time
  - Will you be able to collect and process the sample in order to make it “real time”?
    - Distance of site to analytical lab
  - Do you have diurnal variation?
    - Is an early morning sample representative of water quality at your site?

# Feasibility Studies

- Relationship to health effects
  - *Enterococcus* culture to *Enterococcus* qPCR
    - No need to independently establish a health relationship (NEEAR studies)
  - *E. coli* culture to *E. coli* qPCR
    - May need to independently establish intermediary relationships between culture-based methods
- Result interpretation
  - Numerical and regulatory action decision agreement
    - Environmental/meteorological conditions may influence water quality and method performance
    - Additional information may be needed (sanitary surveys)

# Multiple Lines of Evidence

- Multiple lines of evidence – i.e., multiple tools per site may be necessary in order to:
  - be able to reconcile or evaluate different results from replicates
  - different results from different methods
  - analytical results that seem inconsistent with environmental conditions
- Tools = sanitary surveys, nowcast models, results of multiple analytical methods
- As with any analytical method your results need to make sense!

# Beach Sanitary Surveys

- Guided data gathering at time of sample collection
- Environmental conditions
  - Wave height, turbidity, recent precipitation, presence of algal biomass
- Can be used to predict site specific inhibition
- Can be used in developing predictive models
- Can help you make sense of your data!

# Environmental Data Collected – Routine/Daily Beach Sanitary Survey

- **General Beach Conditions**
  - Air temperature
  - Wind speed/direction
  - Rainfall
  - Weather condition (sunny, etc.)
  - Current speed/direction
  - Wave Height
- **Water Quality**
  - FIB concentrations
  - Water temperature
  - Water color/odor
  - Turbidity (clarity)
- **Bather Load**
  - Total number of people at beach
  - Swimmers/non-swimmers
- **Potential Pollution Sources**
  - Sources of discharge
    - Rivers, outfalls, wetlands, etc.
  - Floatables
  - Amount of debris/litter
  - Amount of algae
    - Stranded on beach
    - Floating/submerged in water
  - Presence of wildlife
    - Gull counts
    - Geese, deer, other
  - Presence of domestic animals
    - Dogs, Horses

# If qPCR is Feasible...

- Choose an appropriate instrument platform
- Familiarize yourself with the method
- Select reagents and controls
- Determine QA/QC procedures
- Provide adequate staff training
- Validate the method
  - Consultation with a qPCR proficient lab recommended
- Conduct side-by-side site specific comparisons of qPCR and culture methods

# Physical Space Requirements

- Segregated work spaces
  - Sample filtration/DNA extraction
  - Preparation of master mix
  - Sample analysis
- Washable surfaces
  - Non-porous
  - UV disinfection, bleach
- Unidirectional workflow
- Dedicated equipment
- Dedicated PPE
- Proper disposal of DNA waste



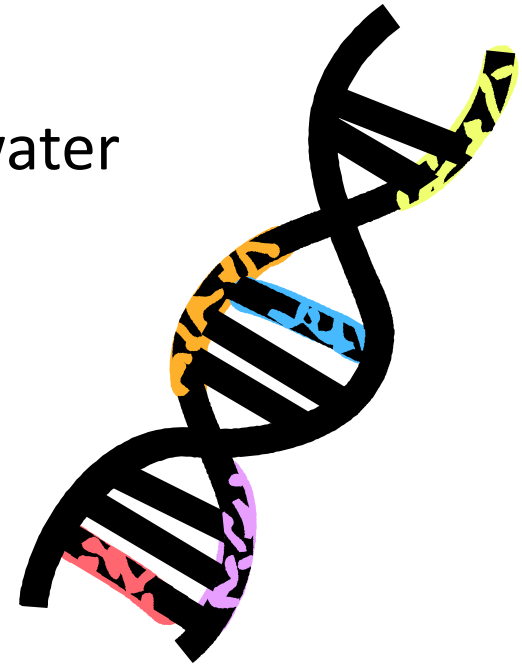
# Choosing an Instrument

- Consider which instrument platform to purchase:
  - Examine your workflow
  - Account for future uses: source tracking, drinking water, wastewater, stormwater, shellfish harvesting, other water research/monitoring?
  - Consumable costs
- Ease of use, software preference
- High throughput or flexibility?
- Balancing variability with cost
  - prepared reagents and standards are more expensive, but will dramatically reduce loss of runs due to errors



# Choice of Molecular Target

- Site Specific Suitability
  - *E. coli* or enterococci for fresh water
  - Enterococci for marine water
  - Bacteroides?
- Wet chemistry or beads
  - Assay time
  - Precision/accuracy
- Sample DNA extract - pre-treatment?
  - Crude
  - Serial Dilution
  - DNA extraction or purification kits



# Sample Handling & Storage

- Storing supplies
  - Calibrators, Lab prepared cells @ -80 °C
  - Controls, SPC @ 4 – 8 °C
  - DNA Reagents, product dependent
  - How long?
    - Stock salmon DNA was good for 5 years @ 4 °C
    - Lab prepared cell suspensions were good for 5 years @ -80 °C
- Storing samples
  - Fresh, analyzed w/i 4 hours of collection
  - Filters, freeze @ -80 °C
  - DNA extracts, good for 7 days @ 4 °C (up to 1% loss/day)
- Mailing samples and/or reagents
  - Degradation can occur if temp is not maintained



# Reagents and Controls

- Reagents
  - Wet Chemistry or bead technology
  - Troubleshooting contamination
- Calibrators and Controls
  - Lab-prepared vs. commercial products
  - Precision (cal curves, replicate sample agreement)
  - Accuracy (calibrators, SPC)
  - Reproducibility (b/w run calibrator agreement)
    - Inter-laboratory validation studies



# QA/QC

- What QC samples should be run?
  - Method Blank (contamination during sample filtration)
  - Filter Blank (contamination in extraction process)
  - NTC (reagent contamination)
  - Calibrator (target quantification check)
  - SPC control (extraction and inhibition control)
- Number/Frequency?
- Definition of inhibition
  - >3 cycle threshold difference b/w cal-SPC and unknown-SPC (EPA definition)

# QA/QC

- Field replicates vs. sample replicates
  - Sample replicates = 2 sub-samples from 1 DNA extraction
- Detection in the negative controls
  - >35 CT in more than a single NTC
  - <45 CT in a third of NTC rxns for a single MM
- Non-agreement/non-consensus
  - Replicate analyses should agree within 1 CT
  - When is it ok to average replicate CT values?
  - How do you know what the right answer is?
- Reporting out “non-detects”
  - Less than half the reciprocal of the dilution factor
  - What does that mean?

# Staff Training Needs

- Aseptic technique!
- Pipetting skills
- Good laboratory practices
- Understanding of molecular concepts
  - Need comprehensive understanding of assay results
- Precision/accuracy
- Reproducibility
- Academic and hands on
  - Learn the basics
  - Observe a trained professional
    - Vendor training
    - National and regional training opportunities
    - Train the trainer (referee labs)
- Data review prior to doing it alone



# Comparative Testing (Method Validation)

- **Within lab**

- Can you continue to use the same procedures/protocols that you used for culture-based assays?
  - Time of sample collection
  - Composite sampling
- Numerical agreement
  - Should/could you expect to see this?
- Regulatory action agreement
  - Will you be as protective to public health and safety?

- **Between labs**

- Inter-laboratory reproducibility studies
- Referee labs

# Method Performance

- Precision/Accuracy/Variability
  - Are the results reproducible?
- Inhibition
  - Do conditions exist which would prevent accurate quantification?
- Comparison to “gold standard”
  - How do the results compare to currently approved methods? To health risk data?
- Do the results make sense?
  - Predictive models, ambient conditions



# The Way Forward

- What is the way forward for broader implementation?
- Further comparative studies
  - Know your beach
- Years of comparative data
- **USE THE SANITARY SURVEY TOOL**
  - Correlations b/w inhibition and ambient conditions
- Virtual Beach
  - Helps make sense of environmental conditions and analytical results

# Points to Consider

- Make sure this analytical method works for you before you start the lab set up process
- Consult with proficient labs
- Exchange sample filters
- Future needs:
  - Sole source or approved QC materials
  - Accreditation
  - Laboratory Inspections
  - Staff Credentials
  - Proficiency Testing

# Acknowledgements

- US EPA BEACH Act
- EPA Contract #EP115000072
- City of Racine, WI
- US EPA Office of Water, NERL, Region 5
- Tamara Anan'eva (qPCR Queen of the WI & EPA OW ORISE Intern)
- A long line of dedicated staff and students:
  - Jennifer Lavender
  - Michelle Leittl
  - Denny Mudd
  - Jenni Creekmur



# Thank You!

